

PERSPECTIVE ARTICLE

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Nanoparticles and vesicles: promising drug-delivery systems

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Abstract

The study of the interaction of drugs with receptors is a subject of great relevance, because it can be applied to the research of new drugs and to the interpretation of the related biological activities. In particular, one of the main challenges that researchers have to face is related to the delivery of the molecule. In the context of healthcare safety, a proper vehicle might indeed improve the targeted and selective absorption of the drug, thus reducing its side effects. Recently, two carriers have gained high attention among the scientific community: nanoparticles and vesicles. The first ones might be obtained from macrocycles such as calixarenes, whose great advantage is the possibility to make their lipophilic structure highly hydrophilic through appropriate functionalizations. The latter arise from phospholipids in natural vesicles or anionic or cationic surfactants in synthetic ones. The present mini-review explores these carriers having possible application in human healthcare.

The study of the interaction of derivatives with human receptors, especially DNA, is an interesting and up-to-date subject, principally for the potential pharmacological activity of DNA-binders. Being the carrier of the genetic information and playing a crucial role in the cellular growth and division, DNA is in fact a biological target of several drugs used for the treatment of various diseases (1). In this context, it is important to outline that different DNAbinding compounds, although interacting with the same target, may exhibit different therapeutic effects like, for example, antimalarial, antitumor and anti-HIV (2–5). Delivering a drug is a key step in the pharmacodynamics of the molecule and this has aroused an increasing interest toward the development of appropriate vehicles. By means of a tailored vehicle, it is in principle possible to improve the healthcare safety for drug treated patients. In this context, vesicles and nanoparticles have been developed as promising new carriers. In the last decades, for example, calixarenes macrocycles have been widely used for the development of new drugdelivery systems, especially nanoparticles. One of the biggest advantage in their use, in fact, is the possibility to make their lipophilic structure hydrophilic through functionalization with several polar groups, such as sulphonates, phosphonates or amines (6). Moreover, due to the presence of two possible sites of functionalization, the lower and the upper rims, calixarenes are also capable to form amphiphilic derivatives (7–10). Several calixarenes have shown both low cytotoxicity, both low immunogenicity (6,11–13): for example, a calix[4]arene phosphonate and a sulphonate one have the same cytotoxicity with respect to that of glucose (12–14).

Vesicles, instead, are self-organized structures constituted by a curved double layer of surfactant molecules, entrapping an inner water pool. In liposomes the monomer units are phospholipids, while in the synthetic vesicles these can be anionic or cationic surfactants. Depending on the preparation method (15-17), it is possible to obtain vesicle dispersions with different characteristics; for example, the formation of unilamellar or multilamellar vesicles can be favoured. An important property of vesicles is the tuneable fluidity of the double layer (18,19). The latter can assume various structures with different permeability and fluidity, depending on temperature. The open double layers and both the uni- or multilamellar vesicles undergo to very well defined structural changes at certain temperatures, called phase transition temperatures (20). The main thermotropic transition phase, interesting for both biological and technological possible applications, is that from the rigid state, commonly called "gel-like", to the fluid state, "liquid crystal-like" phase. It is important to emphasize that by modulating the fluidity of the vesicular nano-aggregate it is possible to modify the solubilisation ability of the phospholipidic double-layer and, consequently, to vary the reactivity of substrates (21-23). Furthermore, DNA is able to interact with liposomes and the supramolecular systems formed are commonly called lipoplexes. The formation of the DNA/Liposome supramolecular structures depends on several parameters, like for example the lipid nature and concentration, the lipid/DNA ratio, the contact time, the temperature and the nature of the reaction medium (24-26). The lipoplexes formed by the association of cationic surfactants and DNA are promising and efficient genic carrier systems (27-30). However, some cationic lipoplexes are also cytotoxic. To get round this difficulty, neutral lipid/DNA systems in the presence of bivalent metal cations have been recently considered (31). These ternary supramolecular systems, analogously to those formed by the cationic liposomes, resulted to be very stable.

In this respect, it is worth mentioning that the supramolecular systems, formed by both the cationic and the neutral liposomes, have been studied only in equilibrium conditions, whereas a few attentions have been devoted to the kinetics of lipoplex formation (32,33). In particular, it has been reported that the lipoplex supramolecular structures are time dependent, and this result puts limits on the transfection efficiency for genic therapy (34). As a consequence, the search of efficient supramolecular structures as alternative vehicles of genic material requires the design of novel systems that can be formed in short time and that are stable, highly compartmentalized and non cytotoxic. In this context, first row transition metal bivalent cations have been used as stabilizing agents of lipoplexes formed by neutral lipid/DNA systems (31). However, while numerous studies have been reported on the interaction of metal complexes with DNA in water solution, to our knowledge in the literature there are not studies on the interaction of metal complexes with DNA confined in nanoscopic biomimetic systems. The only possible exception is a study about the confinement effect in reverse micelles on the intercalation of a copper(II)-Schiff base complex into native DNA (35). The results of this study indicate the occurrence of dramatic structural changes of both the DNA and the metal complex-DNA system, when going from aqueous solution to the micellar phase. Finally, quantum-chemical, molecular mechanics or hybrid computational approaches are useful for assisting the molecular drug design and for studying the nature of the host-guest interactions occurring between the drug and the carrier (36–38).

References

1. Braña MF, Cacho M, Gradillas A, de Pascual-Teresa B, Ramos A. Intercalators as anticancer drugs. Curr Pharm Des. 2001 Nov;7(17):1745–80.

2. Heringova P, Woods J, Mackay FS, Kasparkova J, Sadler PJ, Brabec V. Transplatin is cytotoxic when photoactivated: enhanced formation of DNA cross-links. J Med Chem. 2006 Dec 28;49(26):7792–8.

3. Aloisi GG, Amelia M, Barbafina A, Latterini L, Elisei F, dall' Acqua F, et al. DNA cleavage induced by photoexcited antimalarial drugs: a photophysical and photobiological study. Photochem Photobiol. 2007 Jun;83(3):664–74.

4. Keter FK, Kanyanda S, Lyantagaye SSL, Darkwa J, Rees DJG, Meyer M. In vitro evaluation of dichloro-bis(pyrazole)palladium(II) and dichloro-bis(pyrazole)platinum(II) complexes as anticancer agents. Cancer Chemother Pharmacol. 2008 Mar 19;63(1):127–38.

5. Kalinowska-Lis U, Ochocki J, Matlawska-Wasowska K. Trans geometry in platinum antitumor complexes. Coord Chem Rev. 2008 Jul;252(12–14):1328–45.

6. Perret F, Lazar AN, Coleman AW. Biochemistry of the para-sulfonato-calix[n]arenes. Chem Commun. 2006 Jun 6;(23):2425–38.

7. Shinkai S, Mori S, Koreishi H, Tsubaki T, Manabe O. Hexasulfonated calix[6]arene derivatives: a new class of catalysts, surfactants, and host molecules. J Am Chem Soc. 1986;108(9):2409–16. 8. Shinkai S, Arimura T, Araki K, Kawabata H, Satoh H, Tsubaki T, et al. Syntheses and aggregation properties of new water-soluble calixarenes. J Chem Soc [Perkin 1]. 1989 Jan 1;(11):2039–45.

9. Consoli GML, Granata G, Lo Nigro R, Malandrino G, Geraci C. Spontaneous Self-Assembly of Water-Soluble Nucleotide–Calixarene Conjugates in Small Micelles Coalescing to Microspheres. Langmuir. 2008;24(12):6194–200.

10. Basílio N, Garcia-Rio L. Calixarene-based surfactants: conformational-dependent solvation shells for the alkyl chains. Chemphyschem Eur J Chem Phys Phys Chem. 2012 Jun 18;13(9):2368–76.

Perret F, Coleman AW. Biochemistry of anionic calix[n]arenes. Chem Commun. 2011;47(26):7303.
Martin AD, Houlihan E, Morellini N, Eggers PK, James E, Stubbs KA, et al. Synthesis and Toxicology of p-Phosphonic Acid Calixarenes and O-Alkylated Analogues as Potential Calixarene-Based Phospholipids. ChemPlusChem. 2012;77(4):308–13.

13. Grote Gansey MHB, de Haan AS, Bos ES, Verboom W, Reinhoudt DN. Conjugation, Immunoreactivity, and Immunogenicity of Calix[4]arenes; Model Study to Potential Calix[4] arene-Based Ac3+ Chelators. Bioconjug Chem. 1999;10(4):613–23.

14. Paclet M-H, Rousseau CF, Yannick C, Morel F, Coleman AW. An Absence of Non-specific Immune Response towards para-Sulphonato-calix[n]arenes. J Incl Phenom Macrocycl Chem. 2006 Jun 7;55(3-4):353–7.

15. Bangham AD, Standish MM, Watkins JC. Diffusion of univalent ions across the lamellae of swollen phospholipids. J Mol Biol. 1965 Aug;13(1):238–IN27.

16. Huang C. Studies on phosphatidylcholine vesicles. Formation and physical characteristics. Biochemistry (Mosc). 1969 Jan;8(1):344–52.

17. Szoka F, Papahadjopoulos D. Procedure for preparation of liposomes with large internal aqueous space and high capture by reverse-phase evaporation. Proc Natl Acad Sci. 1978 Sep 1;75(9):4194–8.

18. Nikolova A, Koynova R, Tenchov B, Exerowa D. Chain-melting phase transition in dipalmitoylphosphatidylcholine foam bilayers. Chem Phys Lipids. 1996 Sep 30;83(2):111–21.

19. Gugliotti M, Politi MJ. The role of the gel <=> liquid-crystalline phase transition in the lung surfactant cycle. Biophys Chem. 2001 Feb 15;89(2-3):243–51.

20. Andersson M, Hammarstroem L, Edwards K. Effect of Bilayer Phase Transitions on Vesicle Structure, and its Influence on the Kinetics of Viologen Reduction. J Phys Chem. 1995 Sep 1;99(39):14531–8.

21. Liveri MLT, Sciascia L, Lombardo R, Tesoriere L, Passante E, Livrea MA. Spectrophotometric evidence for the solubilization site of betalain pigments in membrane biomimetic systems. J Agric Food Chem. 2007 Apr 18;55(8):2836–40.

22. Sciascia L, Hauser MJB, Turco Liveri ML. Kinetic evidence for the incorporation of the [(pentamethylcyclopentadienyl) (2,2 -bipyridyl) (aquo)rhodium(III)] complex into DPPC vesicles. Colloids Surf Physicochem Eng Asp. 2008 Jun 5;322(1–3):243–7.

23. Hauser MJB, Müller SC, Sbriziolo C, Liveri MLT. The solubilization site of 5,10,15,20-tetrakis-(2,6-dichlorophenyl)-porphyrin-Mn(III) in DPPC vesicles: A spectrophotometric and tensiometric study. Colloids Surf Physicochem Eng Asp. 2006 Apr 20;278(1–3):212–7.

24. Rejman J, Oberle V, Zuhorn IS, Hoekstra D. Size-dependent internalization of particles via the pathways of clathrin- and caveolae-mediated endocytosis. Biochem J. 2004 Jan 1;377(Pt 1):159–69.

25. Koltover I, Salditt T, Safinya CR. Phase diagram, stability, and overcharging of lamellar cationic lipid-DNA self-assembled complexes. Biophys J. 1999 Aug;77(2):915–24.

26. Heyes JA, Niculescu-Duvaz D, Cooper RG, Springer CJ. Synthesis of Novel Cationic Lipids: Effect of Structural Modification on the Efficiency of Gene Transfer. J Med Chem. 2002 Jan 1;45(1):99–114.

27. Felgner PL, Gadek TR, Holm M, Roman R, Chan HW, Wenz M, et al. Lipofection: a highly efficient, lipid-mediated DNA-transfection procedure. Proc Natl Acad Sci. 1987 Nov 1;84(21):7413–7.

28. Hofland HE, Shephard L, Sullivan SM. Formation of stable cationic lipid/DNA complexes for gene transfer. Proc Natl Acad Sci. 1996 Jul 9;93(14):7305–9.

29. Spector MS, Schnur JM. DNA ordering on a lipid membrane. Science. 1997 Feb 7;275(5301):791–2.

30. Lasic DD, Strey H, Stuart MCA, Podgornik R, Frederik PM. The Structure of DNA–Liposome Complexes. J Am Chem Soc. 1997 Jan 1;119(4):832–3.

31. Francescangeli O, Stanic V, Gobbi L, Bruni P, lacussi M, Tosi G, et al. Structure of self-assembled liposome-DNA-metal complexes. Phys Rev E. 2003 Jan 16;67(1):011904.

32. Barreleiro PCA, Lindman B. The Kinetics of DNA–Cationic Vesicle Complex Formation. J Phys Chem B. 2003 Jun 1;107(25):6208–13.

33. Elkady A, Tychinsky VP, Vyshenskaja TV, Sebyakin YL, Yaminskii IV, Zhdanov RI, et al. Laser Contrast and Atomic Force Microscopy of Complexes between Plasmid DNA and a New Dicationic Lipid. Dokl Biochem Biophys. 2004 May 1;396(1-6):161–4.

34. Süleymanoglu E. Adiabatic differential

scanning calorimetric study of divalent cation induced DNA - DPPC liposome formulation compacted for gene delivery. Braz Arch Biol Technol. 2004 Nov;47(6):881–5.

35. Barone G, Longo A, Ruggirello A, Silvestri A, Terenzi A, Turco Liveri V. Confinement effects on the interaction of native DNA with Cu(II)-5-(triethylammoniummethyl)salicylidene orthophenylendiiminate in $C_{12}E_4$ liquid crystals. Dalton Trans. 2008;(31):4172–8.

36. Zielenkiewicz W, Marcinowicz A, Poznański J, Cherenok S, Kalchenko V. Complexation of isoleucine by phosphorylated calix[4]arene in methanol followed by calorimetry, NMR and UV–VIS spectroscopies, and molecular modeling methods. J. Mol. Liq. 2005 Jul 30;121(1):8–14.

37. Sangpheak W, Khuntawee W, Wolschann P, Pongsawasdi P, Rungrotmongkol T. Enhanced stability of a naringenin/2,6-dimethyl β-cyclodextrin inclusion complex: Molecular dynamics and free energy calculations based on MM- and QM-PBSA/GBSA. J Mol. Graph. Model. 2014 May;50:10–5.

38. Macháčková M, Tokarský J, Čapková P. A simple molecular modeling method for the characterization of polymeric drug carriers. Eur. J. Pharm. Sci. 2013 Jan 23;48(1–2):316–22.